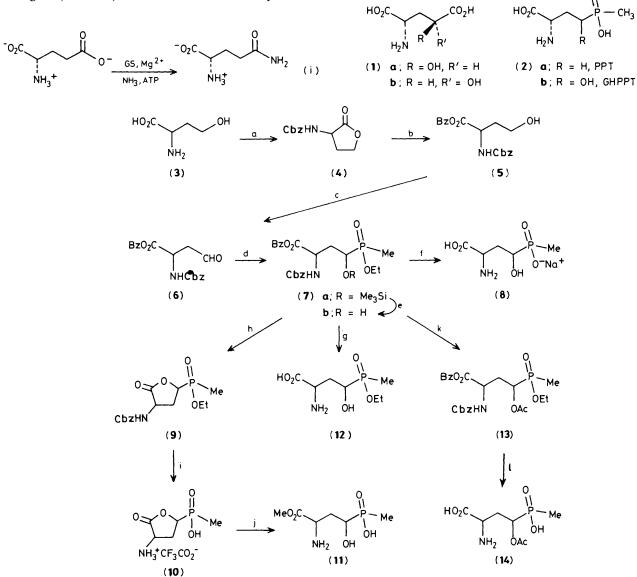
Synthesis of D,L-γ-Hydroxyphosphinothricin, a Potent New Inhibitor of Glutamine Synthetase

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A flexible synthesis of D,L-Y-hydroxyphosphinothricin (GHPPT), a potent inhibitor of the enzyme glutamine synthetase, is described which features silicon-mediated addition of ethyl methylphosphinate to benzyl 2-benzyloxy-carbonylamino-4-oxobutyrate; selective deprotections yield the title compound and various derivatives.

The enzyme glutamine synthetase (GS; E.C. 6.3.1.2) catalyses an important reaction in nitrogen metabolism, the conversion of L-glutamate into L-glutamine [reaction (i)].¹ While L-glutamic acid serves as the natural substrate for the enzyme, it was reported some time ago that the γ -hydroxy analogues (**1a** and **b**) also function as remarkably efficient substrates, the *threo*-isomer (1a) possessing a lower K_m value than that of L-glutamic acid itself.^{2,3} We have been engaged in the design of analogues of phosphinothricin (PPT) (2a), a naturally occurring phosphinic acid mimic of L-glutamic acid^{4,5}



Scheme 1. *Reagents*: a, CbzCl, H₂O, NaHCO₃; 100 °C (0.1 mm Hg), 83%; b, NaOH, MeOH; PhCH₂Br, Me₂NCHO, 23 °C, 24 h, 80%; c, Me₂SO, CH₂Cl₂, (COCl)₂, -78 °C; Et₃N, 100%; d, MeP(O)HOEt, BSA, CH₂Cl₂, 23 °C, 90%; e, HF, H₂O, MeCN, 95%; f, Me₃SiBr, CH₂Cl₂, 23 °C; H₂, 10% Pd/C, H₂O, MeOH, 1 equiv. NaOH, 55%; g, H₂, 10% Pd/C, H₂O, EtOH, 55%; h, CF₃CO₂H, CH₂Cl₂, 23 °C, 71% from (7**a**) or 100% from (7**b**); i, Me₃SiBr, CH₂Cl₂, 23 °C; H₂, 10% Pd/C, H₂O, MeOH; CF₃CO₂H, 55 °C, 98%; j, MeOH, reflux, 50%; k, Ac₂O, 4-*N*,*N*-dimethylaminopyridine, pyridine, 95%; l, Me₃SiBr, CH₂Cl₂, 23 °C; H₂, 10% Pd/C, H₂O, EtOH, 47%.

and an effective herbicide.^{6,7} It was of interest to investigate the effect of introducing substituents found in alternative GS substrates into the structure of transition state analogue inhibitors such as (2a).⁸ We report here an efficient synthesis of D,L- γ -hydroxyphosphinothricin (GHPPT), a novel phosphinothricin analogue which functions as a potent competitive inhibitor of glutamine synthetase.

A flexible route was developed which also afforded selectively functionalized derivatives of interest in probing structure-activity relationships in GS inhibitors. D,Lhomoserine (3) was converted via the lactone $(4)^9$ into the N-Cbz benzyl ester (5) in 66% yield. Previously described methods for oxidizing (5) to benzyl 2-benzyloxycarbonylamino-4-oxobutyrate (6) proved unsatisfactory;10 however, Swern conditions (oxalyl chloride, Me₂SO in CH₂Cl₂ at -78 °C followed by triethylamine) were found to yield (6) almost quantitatively, regardless of scale. Addition of ethyl methylphosphinate to (6) in the presence of bis(trimethylsilvl)acetamide(BSA)¹¹ smoothly gave the silvl ether (7a) and the alcohol (7b) in 63 and 27% yields, respectively, after silica gel chromatography (EtOAc/hexane, followed by EtOAc/ $Pr^{i}OH$). The ether (7a) and the alcohol (7b) were both obtained as mixtures of four chromatographically inseparable diastereoisomers. Treatment of (7a) with bromotrimethylsilane in methylene chloride, followed by hydrogenation in aqueous methanol containing an equivalent of sodium hydroxide, filtration through Celite, and precipitation with acetone afforded the sodium salt of D,L-Y-hydroxyphosphinothricin (GHPPT) (8) in 55% yield. It was obtained as a hygroscopic solid, m.p. 100-120 °C, consisting of a 56:44 mixture of diastereoisomers.[†]

The alcohol (7b) served as a key intermediate for the

(10), m.p. 110–115 °C (decomp.); ¹H n.m.r. [400 MHz; (CD₃)₂SO; 60: 40 diastereoisomeric mixture] δ 1.31* (d, 3H, ²J_{CP} 14.0 Hz), 1.32 (d, ²J_{CP} 14.9 Hz), 2.30–2.46* (m, 1H), 2.66* (m, 0.6 H), 2.79 (m, 0.4 H), 4.37* (m, 1H), 4.61* (t, 0.6 H, J7.6 Hz), and 4.76 (dd, 0.4 H, J4.0 and 9.3 Hz); ¹³C n.m.r. [100.6 MHz; (CD₃)₂SO] δ 14.84* (d, ¹J_{CP} 96.5 Hz), 28.58, 29.33*, 50.46, 51.37* (d, ³J_{CP} 4.7 Hz), 77.62* (d, ¹J_{CP} 109.8 Hz), 78.34 (d, ¹J_{CP} 106.6 Hz), 118.80* (q, ¹J_{CF} 291.6 Hz), 165.28* (q, ²J_{CF} 35.4 Hz), 175.91*, and 175.98; ³¹P n.m.r. [40.3 MHz, (CD₃)₂SO] δ 35.47* and 37.01.

(14) m.p. 124—126 °C (decomp.) ¹H n.m.r. (400 MHz; D₂O; 90 : 10 diastereoisomeric mixture) δ 1.27 (d, ²J_{CP} 14.5 Hz), 1.28* (d, 3H, ²J_{CP} 14.4), 2.15 (s), 2.17* (s, 3H), 2.28—2.50* (m, 2H), 4.02* (dd, 1H, J 4.9 and 9.1 Hz), 4.12 (t, J 5.9 Hz), 5.05—5.13* (m, 1H), and 5.48 (m); ¹³C n.m.r. (75.4 MHz; D₂O; major isomer only) δ 15.39 (d, ¹J_{CP} 95.6 Hz), 23.04, 32.96, 53.64 (d, ³J_{CP} 9.1 Hz), 71.96 (d, ¹J_{CP} 107.1 Hz), 174.42, and 176.16 (d, ³J_{CP} 4.3 Hz); ³¹P n.m.r. (40.3 MHz, D₂O) δ 37.62.

derivatives (14) (GAPPT), (10), (11), and (12) (see Scheme 1).

Under standard assay conditions[‡] with bacterial (*E. coli*) glutamine synthetase,[§] in which the observed substrate K_m value for glutamate was 3.3 mM, a value of 1.6 μ M was obtained for K_i of the inhibitor (8) (GHPPT), one of the most potent binding constants yet seen for inhibition of GS under reversible conditions.^{4,5} A K_i value of 33 μ M was determined for (14) (GAPPT). Both compounds were comparably effective inhibitors of GS from sheep brain and sorghum leaf. The compounds (8) (GHPPT), (14) (GAPPT), (10), and (11) all showed strong herbicidal activity against a broad range of plant species. Interestingly, the phosphinate ester (12) was completely inactive, supporting the hypothesis that enzyme mediated phosphorylation of the phosphinic acid moiety by ATP is required for inhibition.⁵ Biochemical studies in this series will be described in greater detail elsewhere.

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References

- 1 E. R. Stadtman and A. Ginsberg, 'The Enzymes,' ed. P. D. Boyer, Academic Press, New York, 1974, vol. 10, p. 755.
- 2 H. M. Kagan and A. Meister, *Biochemistry*, 1966, **5**, 2423; substrate K_m values (mm) of 3.9, 2.4, and 5.6 were obtained for glutamate, *threo*- γ -hydroxyglutamate, and *erythro*- γ -hydroxyglutamate.
- 3 Recent work describes these compounds as substrates of *Chlorella* GS: N. A. Firsova, L. V. Alekseeva, K. M. Selivanova, and Z. G. Evstigneeva, *Biokhimiya*, 1986, 51, 980 (*Chem. Abstr.*, 1986, 105, 129897p).
- 4 E. Bayer, K. H. Gugel, K. Hägele, H. Hagenmaier, S. Jessipow, W. A. König, and H. Zähner, *Helv. Chim. Acta*, 1972, 55, 224.
- 5 J. A. Colanduoni and J. J. Villafranca, *Bioorg. Chem.*, 1986, 14, 163.
- 6 S. M. Ridley and S. F. McNally, Plant Sci., 1985, 39, 31.
- 7 E. W. Logusch, Tetrahedron Lett., 1986, 27, 5935.
- 8 For a study of substrate-inhibitor correlations for the enzyme thermolysin, see P. A. Bartlett and C. K. Marlowe, *Biochemistry*, 1983, **22**, 4618.
- 9 M. Flavin and C. Slaughter, J. Biol. Chem., 1960, 235, 1103.
- 10 Previous preparations of D,L-(6): C.-D. Chang and J. K. Coward, J. Med. Chem., 1976, 19, 684; J. M. Domagala and T. H. Haskell, J. Org. Chem., 1981, 46, 134; L-(6): D. D. Keith, J. A. Tortora, K. Ineichen, and W. Leimgruber, Tetrahedron, 1975, 31, 2633; G. A. Mock and J. G. Moffatt, Nucleic Acids Res., 1982, 10, 6223.
- J. K. Thottathil, D. E. Ryono, C. A. Przybyla, J. L. Moniot, and R. Neubeck, *Tetrahedron Lett.*, 1984, 25, 4741; D. A. Evans, K. M. Hurst, and J. M. Takacs, *J. Am. Chem. Soc.*, 1978, 100, 3467. For a base-catalysed addition of a phosphite to a protected aspartic acid β-semialdehyde, see M. Kamber and G. Just, *Can. J. Chem.*, 1985, 63, 823.

 \ddagger Values of K_i were determined by the Lineweaver-Burke method, utilizing data obtained from assays measuring either phosphate or ADP release under conditions in which irreversible inhibition was not significant.

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[†] All compounds isolated gave satisfactory spectroscopic (¹H and ³¹P n.m.r.) and analytical data. Selected physical data include (spectral resonances corresponding to the major diastereoisomer in isomeric mixtures are denoted with an asterisk): (8), m.p. 100–120 °C (gradual decomp.); ¹H n.m.r. (300 MHz; D₂O) δ 1.25 (d, 3H, ²J_{CP} 14.2 Hz), 1.90–2.35 (series of m, 2H), and 3.69–3.93 (m, 2H); ¹³C n.m.r.(90.6 MHz; D₂O; 56:44 diastereomeric mixture) δ 12.78* (d, ¹J_{CP} 91.9 Hz), 32.09*, 32.35, 53.59* (d, ³J_{CP} 11.8 Hz), 54.30 (d, ³J_{CP} 11.6 Hz), 68.62* (d, ¹J_{CP} 110.2 Hz), 69.65 (d, ¹J_{CP} 109.7 Hz), and 175.86, 176.10*; ³¹P n.m.r. (40.3 MHz; D₂O) δ 40.47.